

Non Linear Dynamics and Gene Networks

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Non-linearity means that the dynamical behavior of the system cannot be viewed as a superposition of the elementary effects of its components, nor reconstructed from elementary "modes".

In simple terms, doubling the input does not necessarily double the output.

Ricardo LIMA

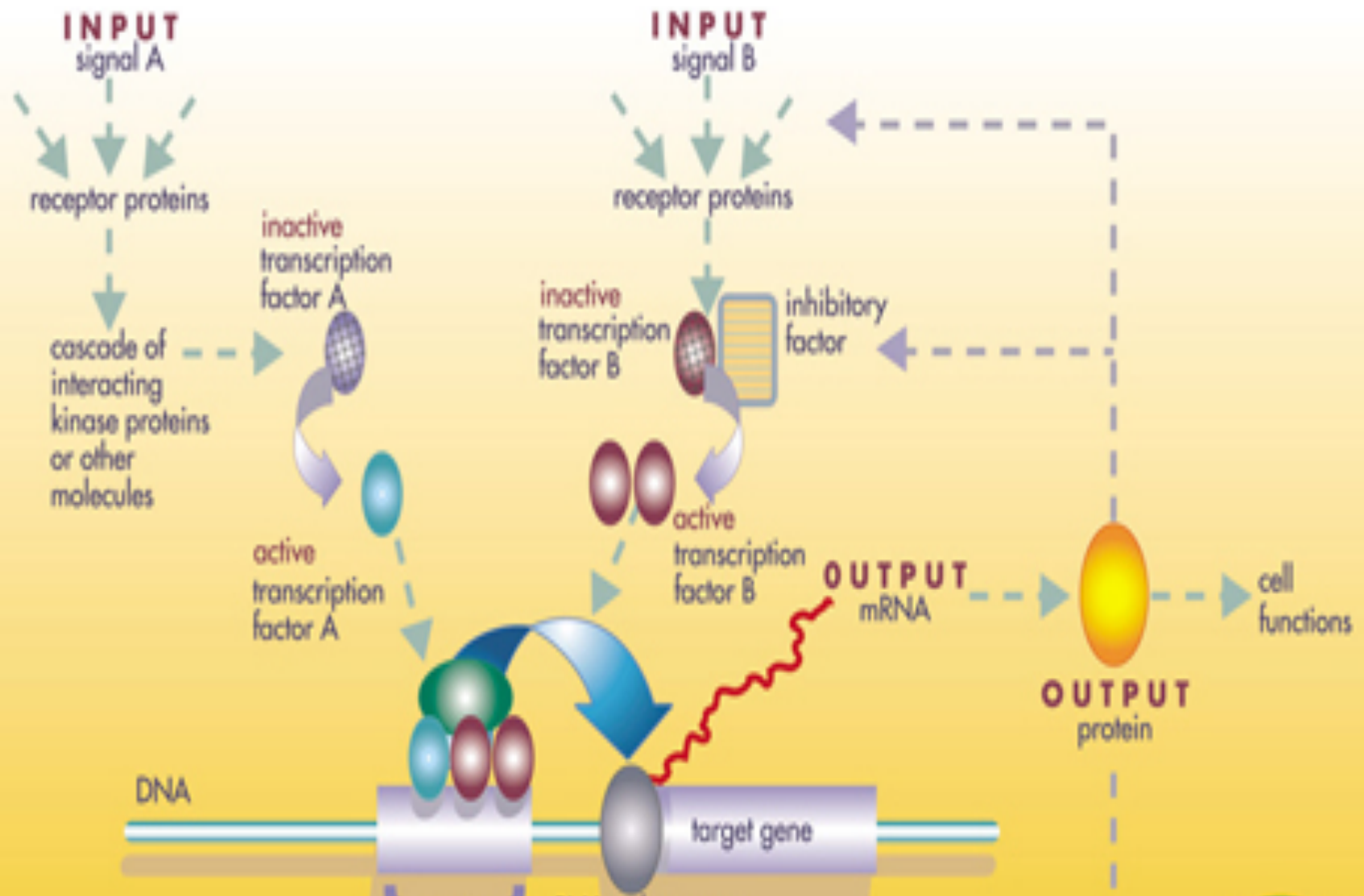
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Research, advise, education : a freelance project

- 1. GENE NETWORK = INTERACTION GRAPH
- 2. DYNAMICS ON THE CONFIGURATION SET
- 3. EXAMPLES
- 4. SOME OPEN QUESTIONS
- 5. SUPPLEMENTARY POINTS
- 6. FINAL QUESTIONS

A GENE REGULATORY NETWORK



transcription factors

of eukaryotic cells

1 Activator proteins bind to pieces of DNA called enhancers. Their binding causes the DNA to bend, bringing them near a gene promoter, even though they may be thousands of base pairs away.

Enhancers

Activator proteins

Other transcription factor proteins

2 Other transcription factor proteins join the activator proteins, forming a protein complex which binds to the gene promoter.

Gene

Promoter

3 This protein complex makes it easier for RNA polymerase to attach to the promoter and start transcribing a gene.

RNA polymerase

note

This diagram simplifies the DNA greatly—promoters, enhancers, and insulators can be dozens or even hundreds of base pairs long.

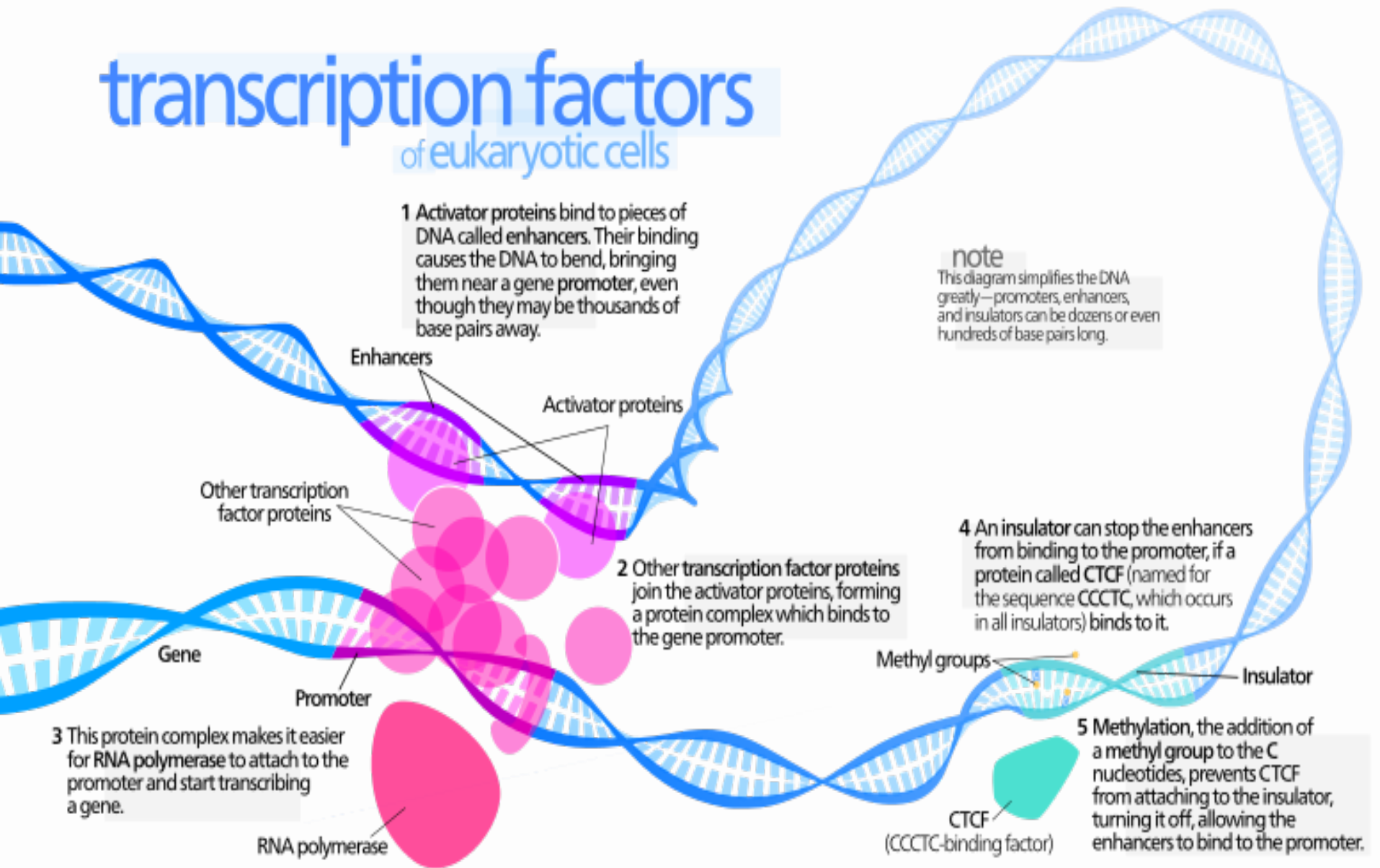
4 An insulator can stop the enhancers from binding to the promoter, if a protein called CTCF (named for the sequence CCCTC, which occurs in all insulators) binds to it.

Methyl groups

Insulator

5 Methylation, the addition of a methyl group to the C nucleotides, prevents CTCF from attaching to the insulator, turning it off, allowing the enhancers to bind to the promoter.

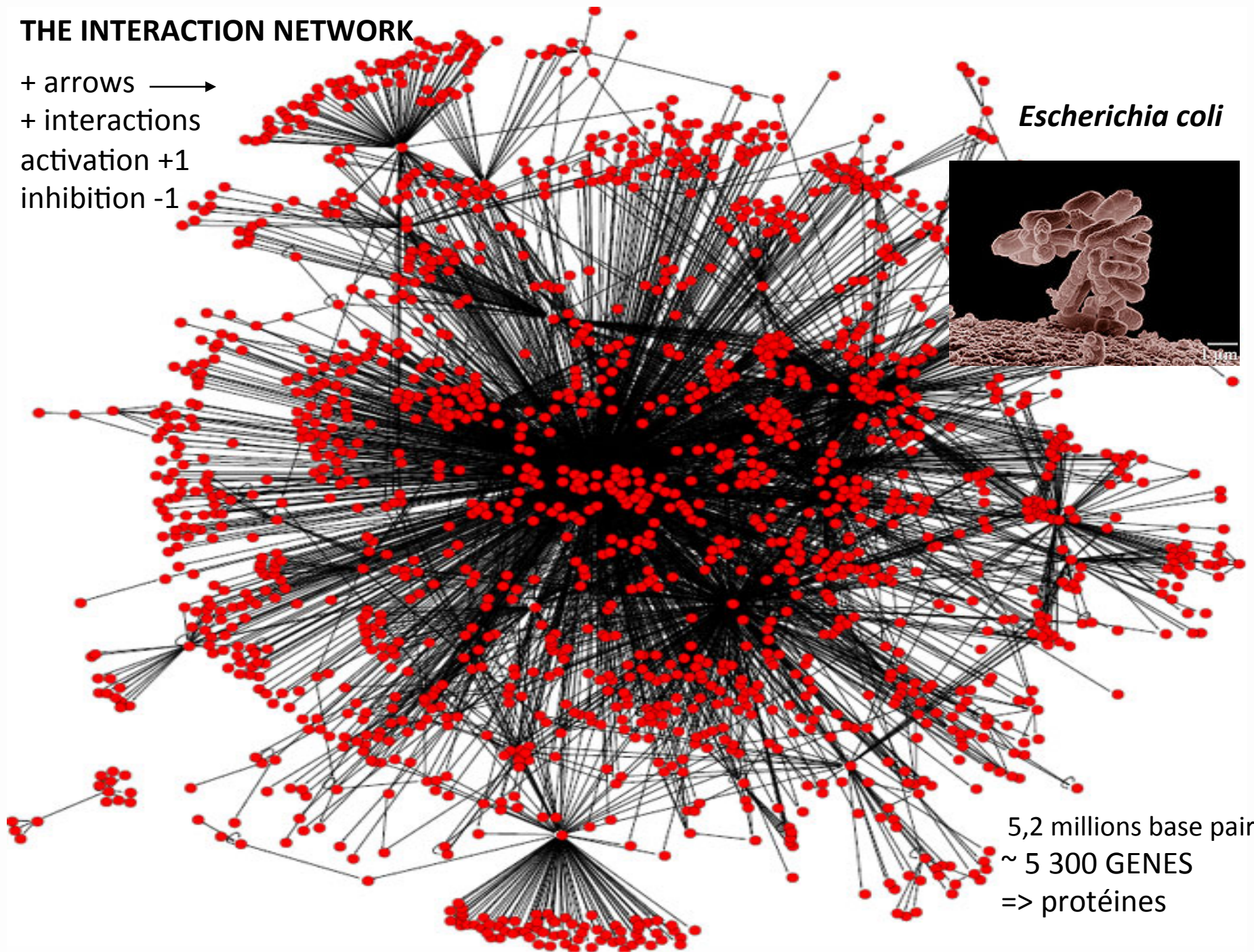
CTCF
(CCCTC-binding factor)



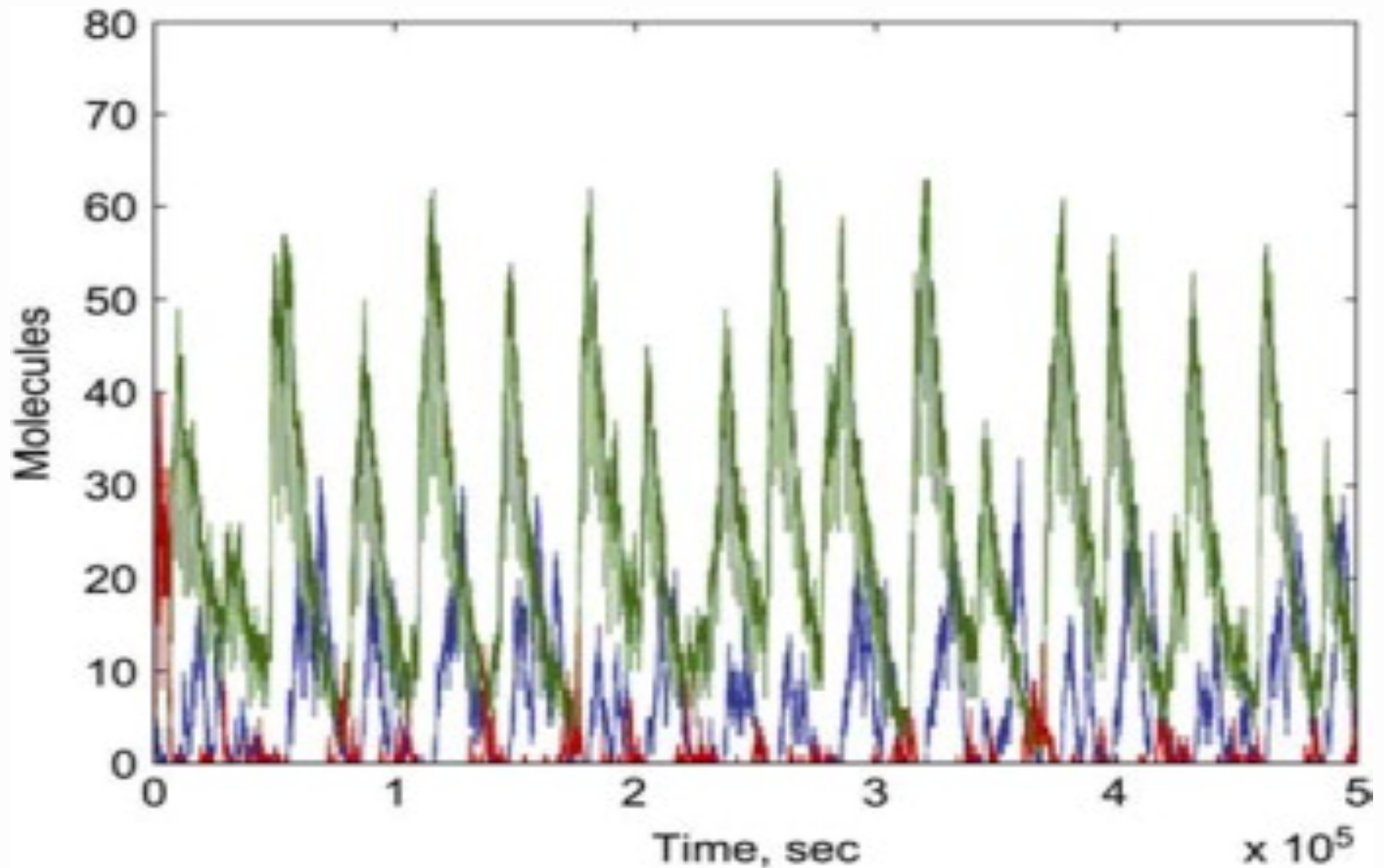
THE INTERACTION NETWORK

+ arrows →
+ interactions
activation +1
inhibition -1

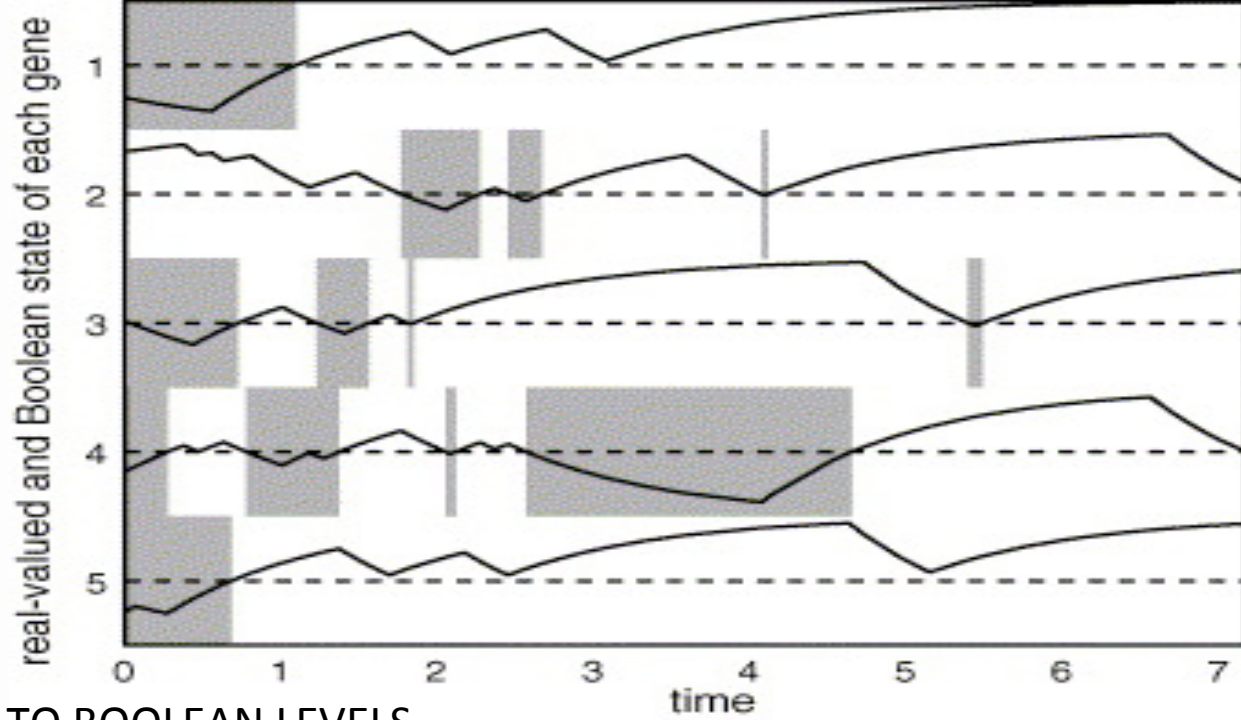
Escherichia coli



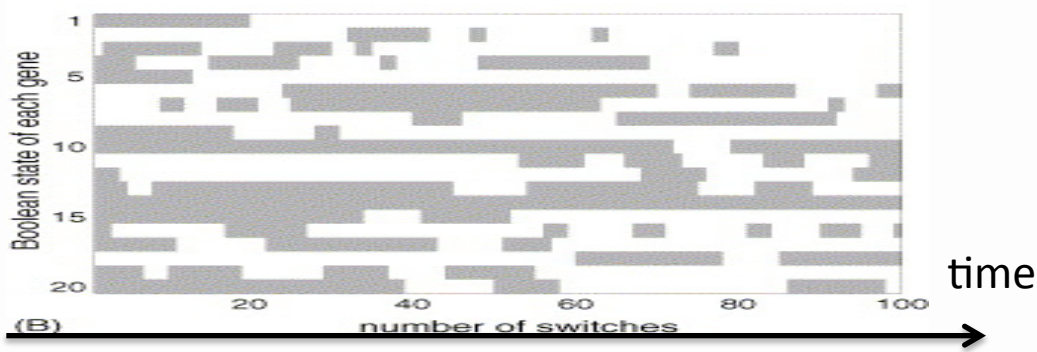
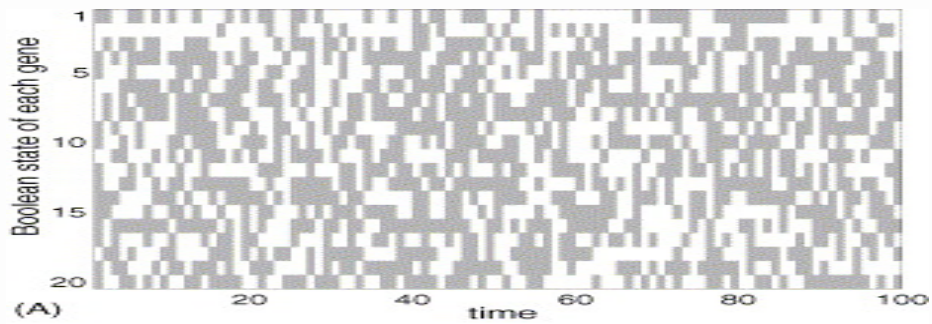
5,2 millions base pairs
~ 5 300 GENES
=> protéines



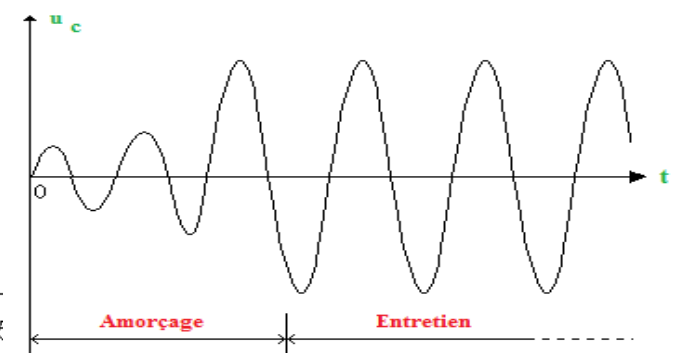
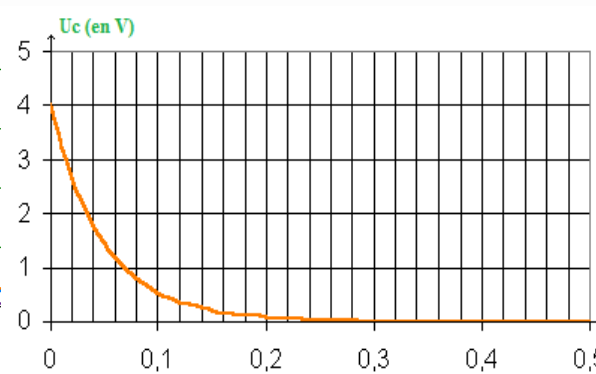
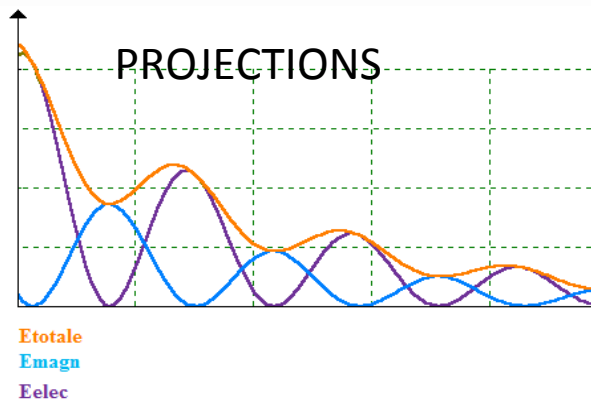
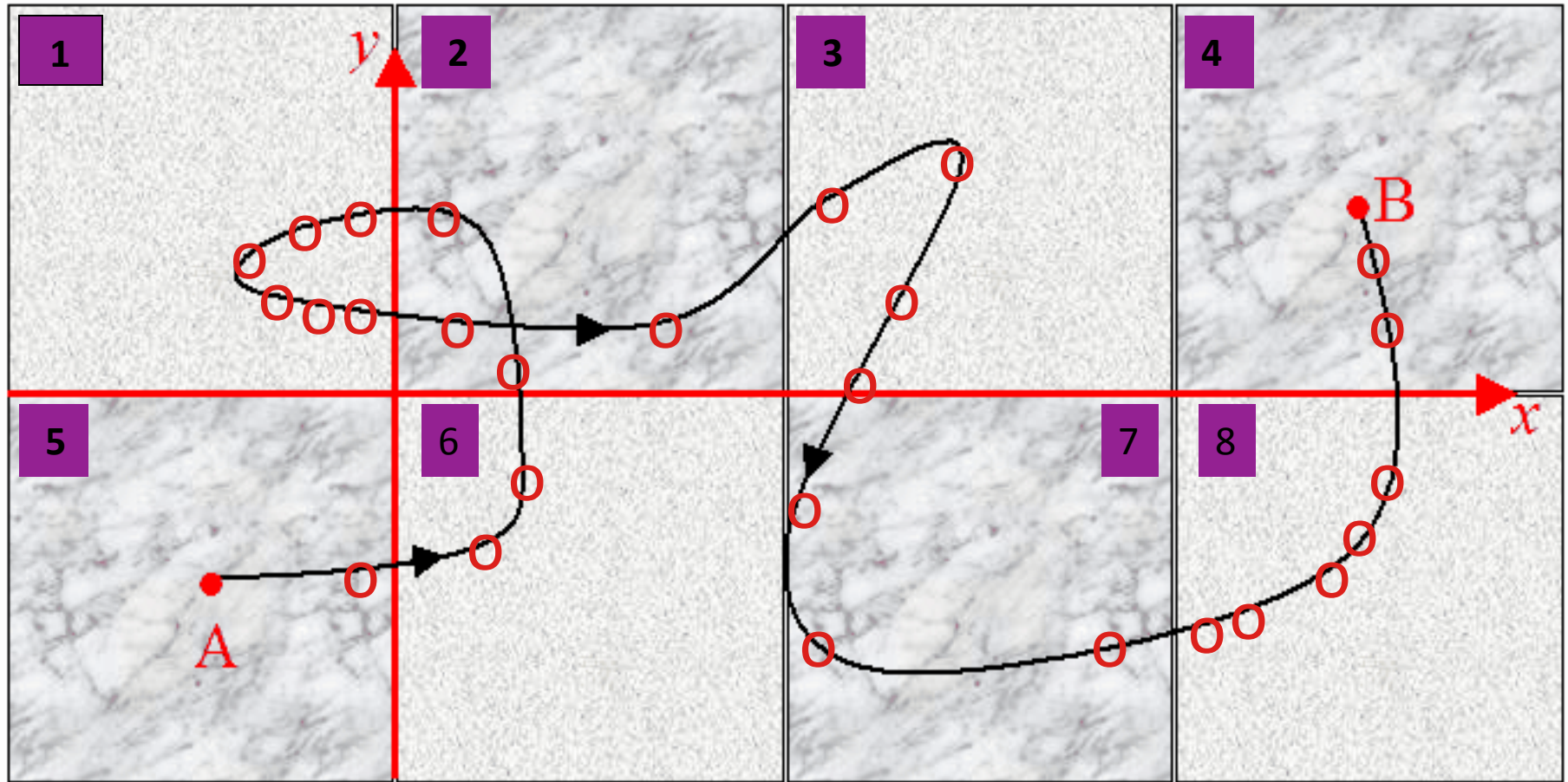
**Dynamical behavior:
Dlac (red), Dtet (blue) and Dara (green) proteins.**



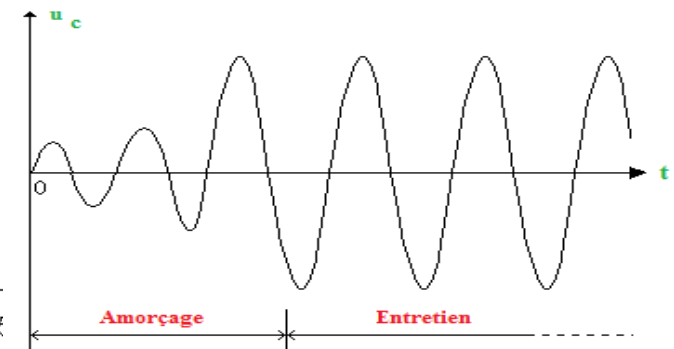
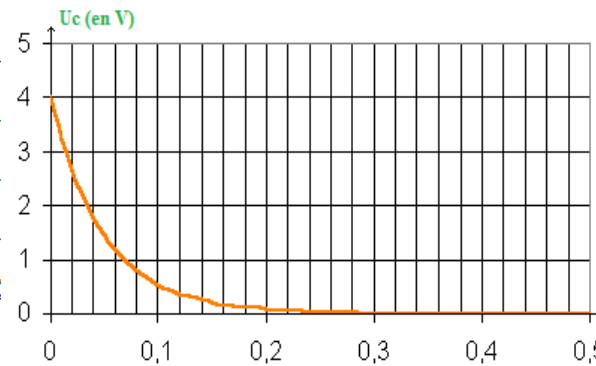
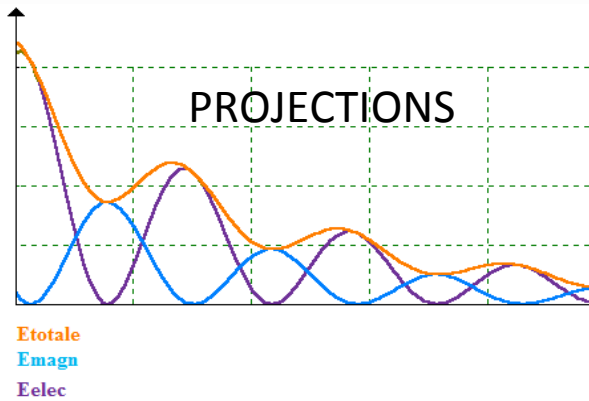
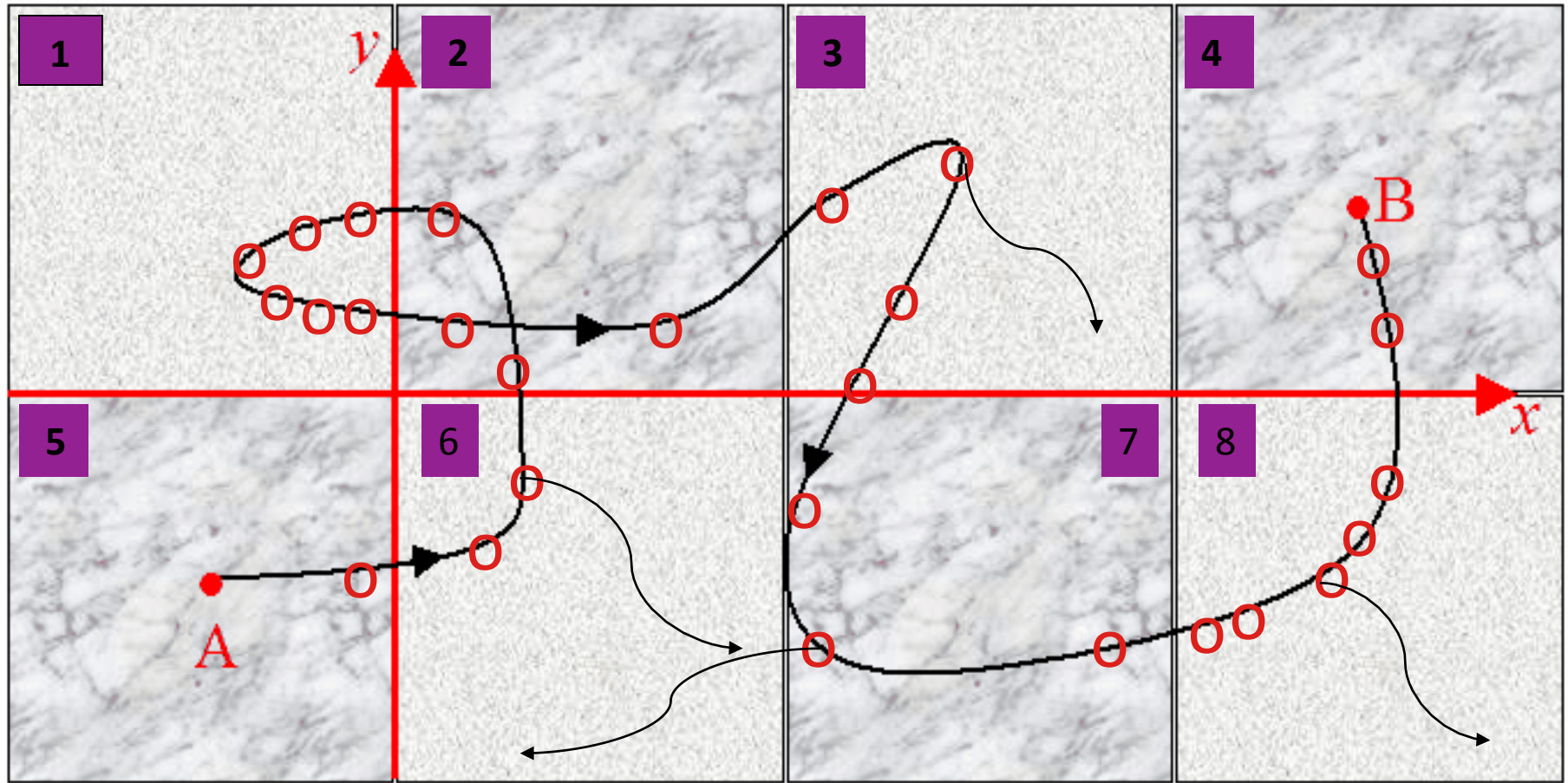
FROM REAL TO BOOLEAN LEVELS



Configuration Space: DETERMINISTIC DYNAMICS

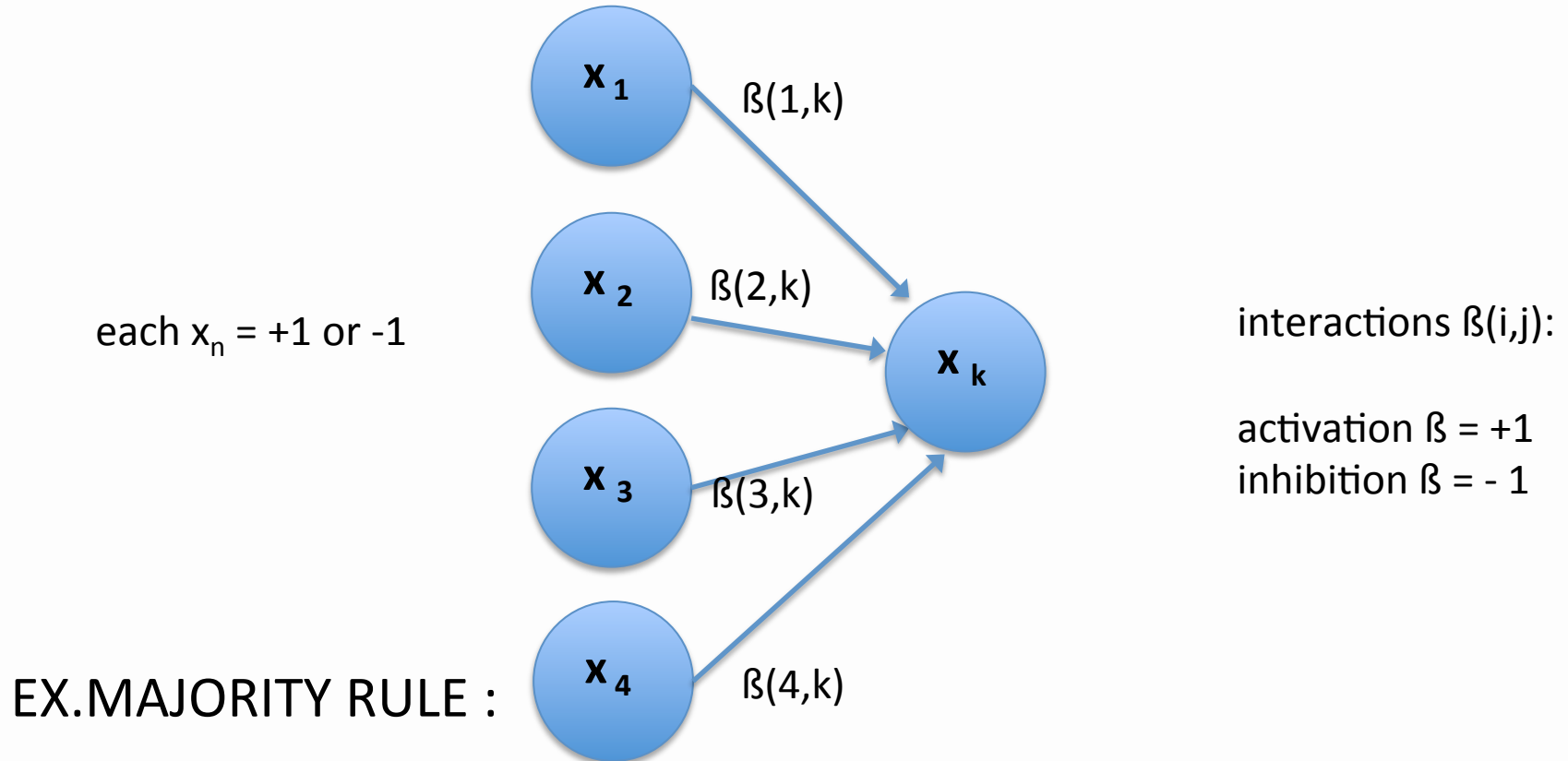


Configuration Space: MARKOV DYNAMICS






DYNAMICS ON THE CONFIGURATION SPACE



at time t : CONFIGURATION $x_1(t), x_2(t), x_3(t), x_4(t), \dots x_k(t) \dots$

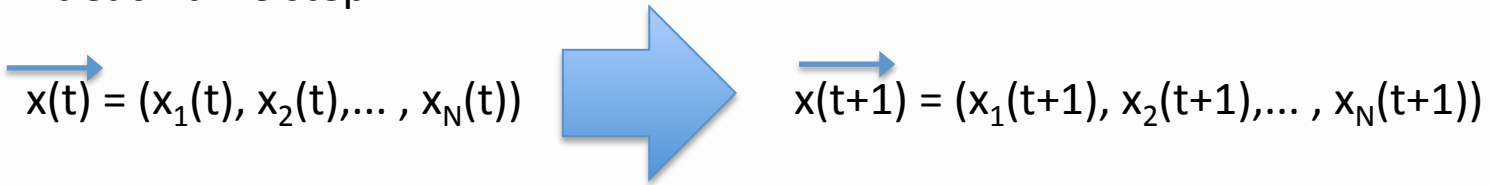
 $x_k(t+1) = +1$ if $\sum = \beta(1,k) x_1(t) + \beta(2,k) x_2(t) + \beta(3,k) x_3(t) + \beta(4,k) x_4(t) > 0$

$x_k(t+1) = +1$ with probability p_k or -1 with probability $(1-p_k)$ if $\sum = 0$

$x_k(t+1) = -1$ if $\sum = \beta(1,k) x_1(t) + \beta(2,k) x_2(t) + \beta(3,k) x_3(t) + \beta(4,k) x_4(t) < 0$

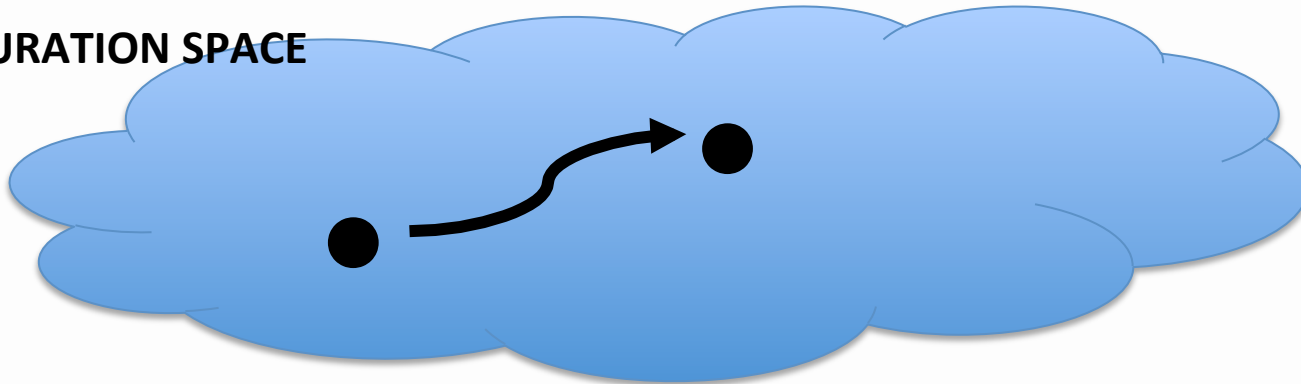
FROM THE INTERACTION NETWORK TO THE DYNAMICS

At each time step:



CONFIGURATION SPACE

Ω

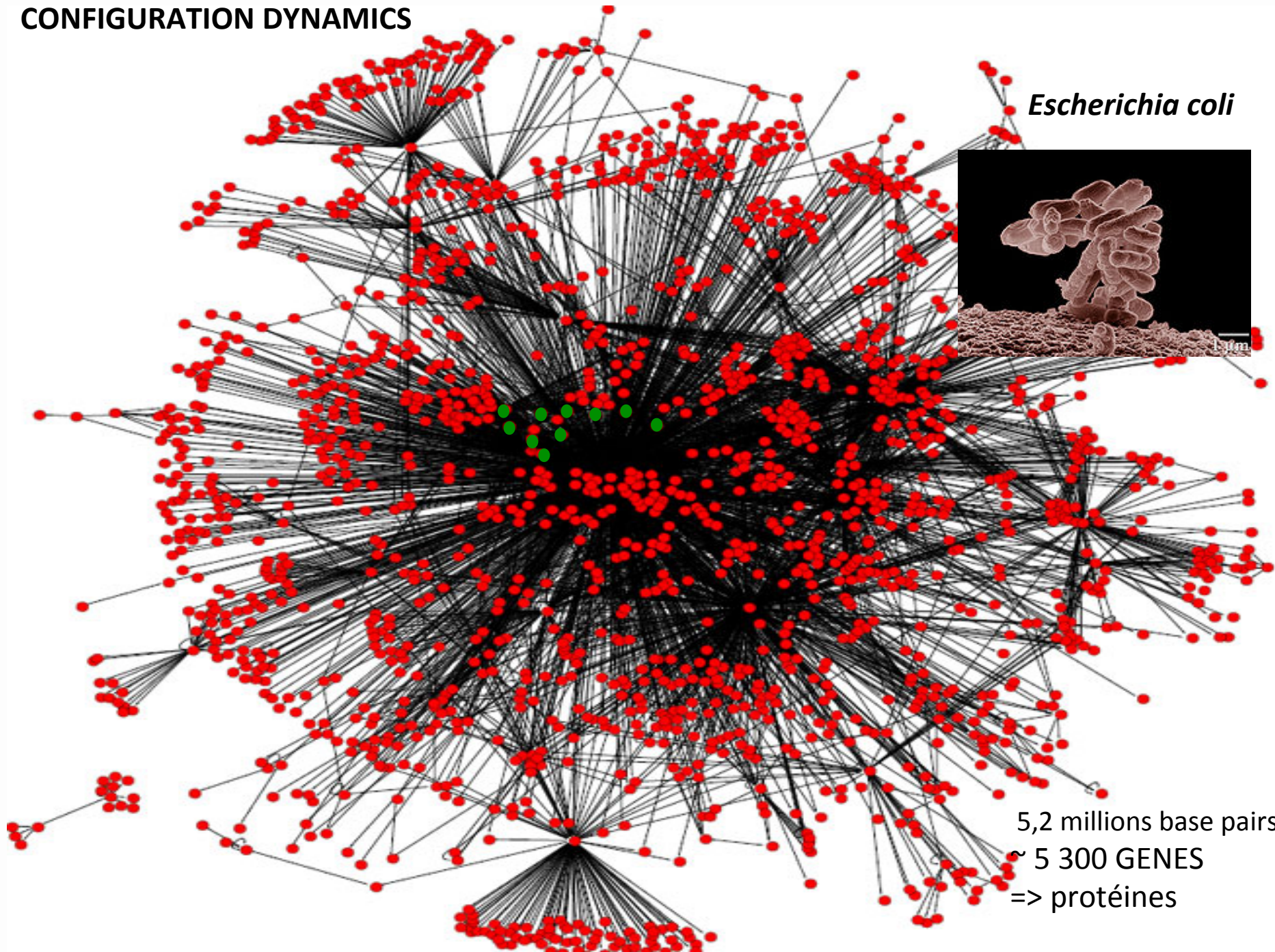


The **INTERACTION NETWORK** defines the **DYNAMICAL RULE** on Ω

Determinist X Probabilist

CONFIGURATION DYNAMICS

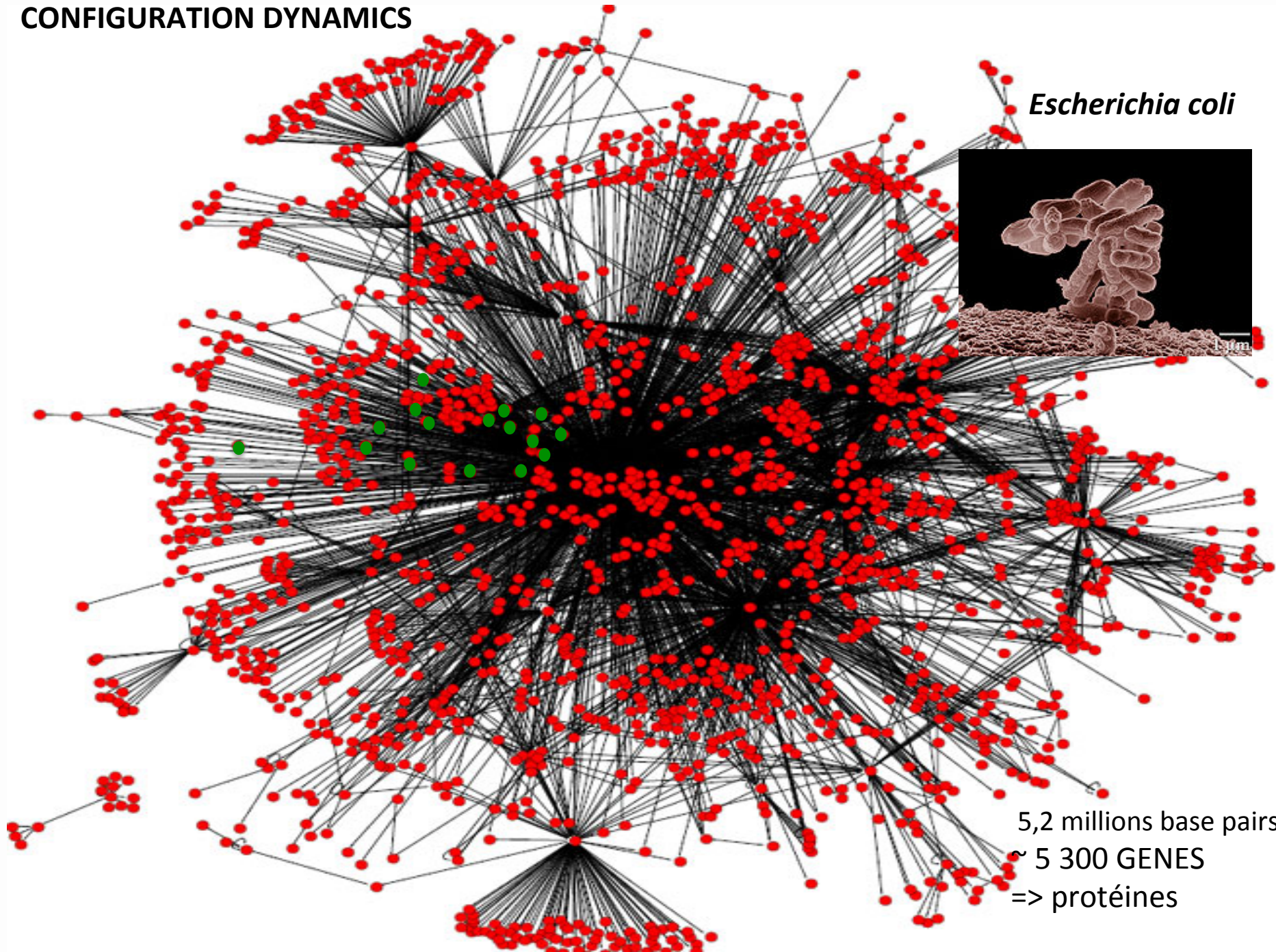
Escherichia coli



5,2 millions base pairs
~ 5 300 GENES
=> protéines

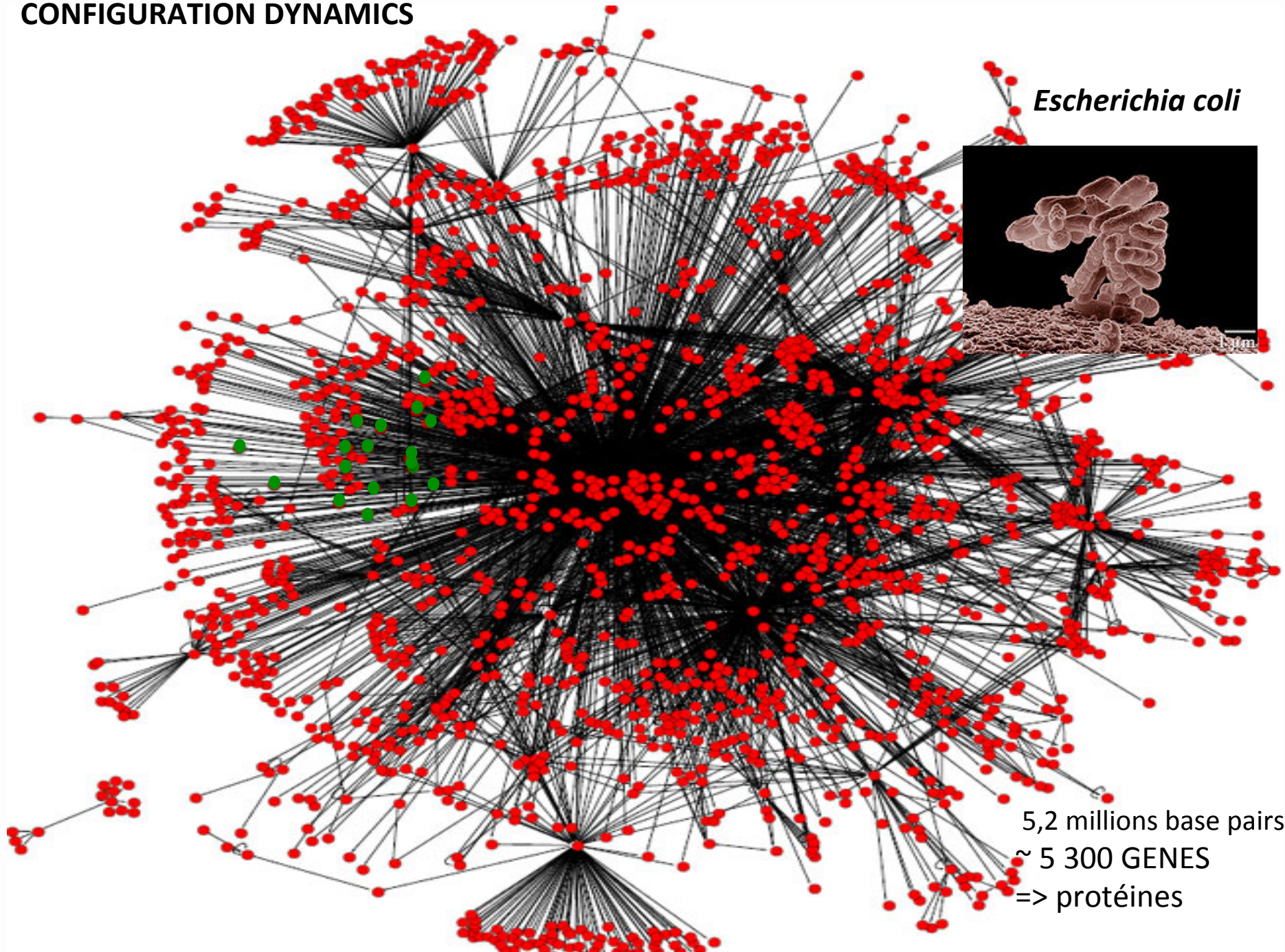
CONFIGURATION DYNAMICS

Escherichia coli

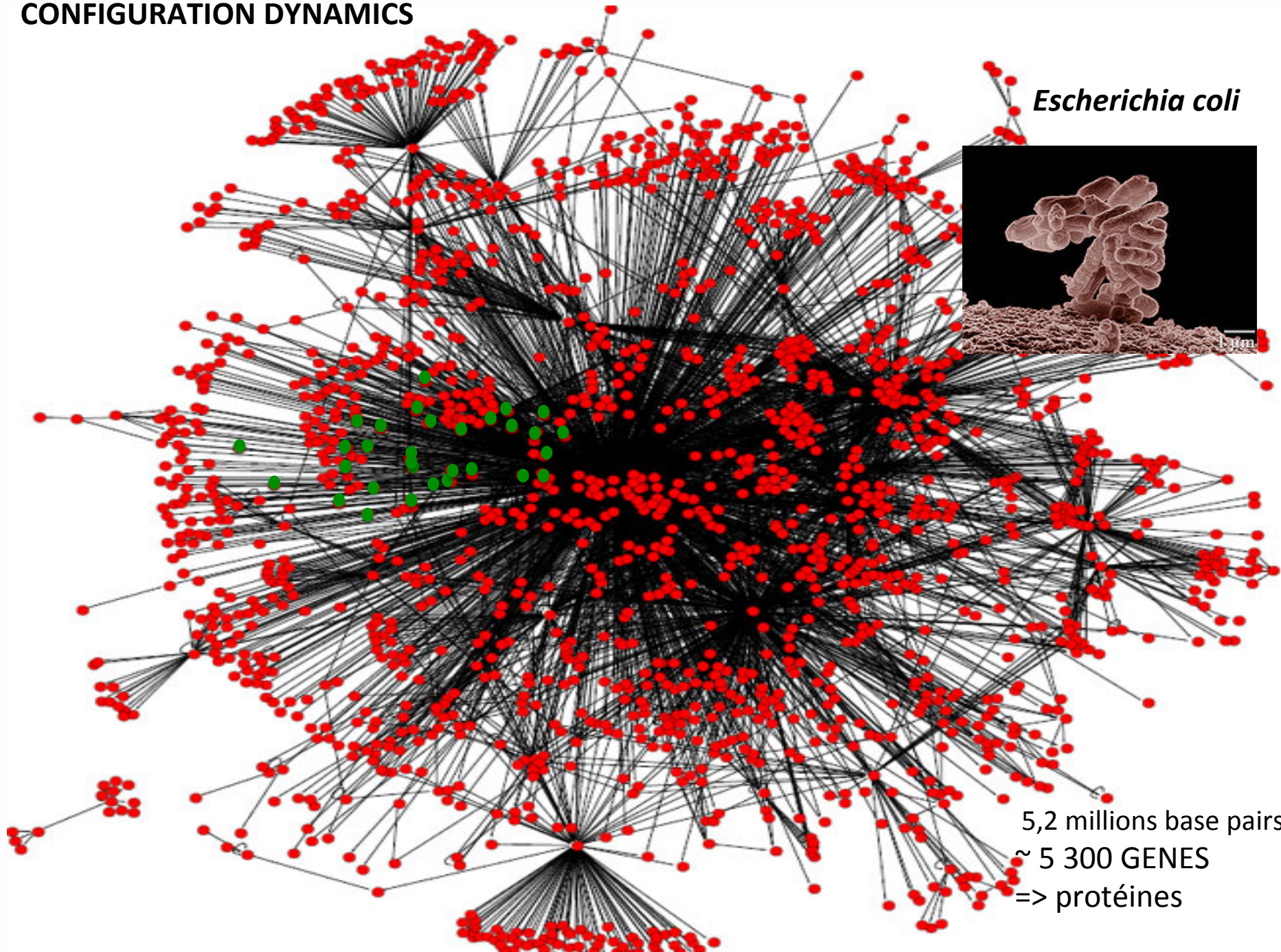


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CONFIGURATION DYNAMICS



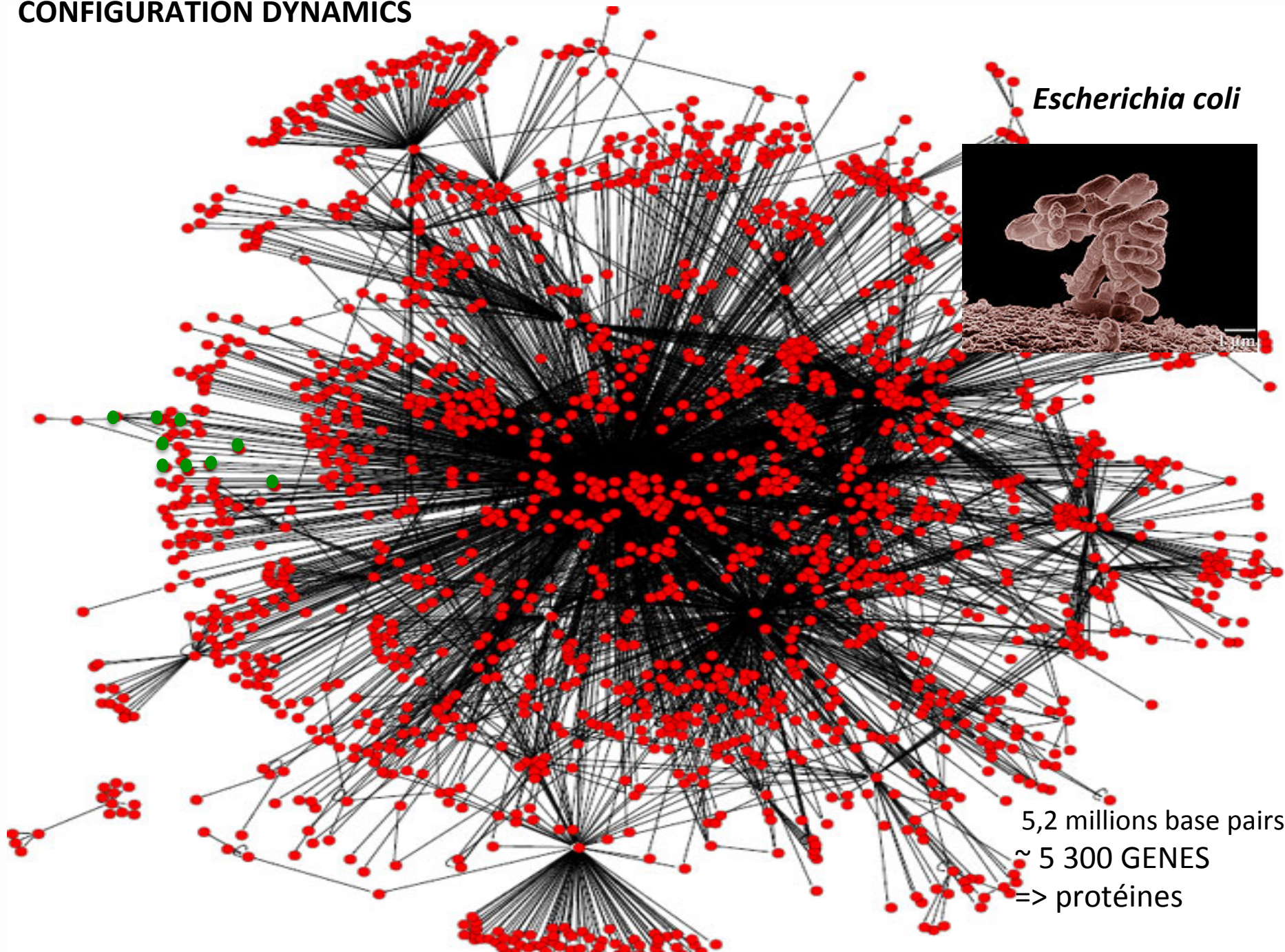
CONFIGURATION DYNAMICS



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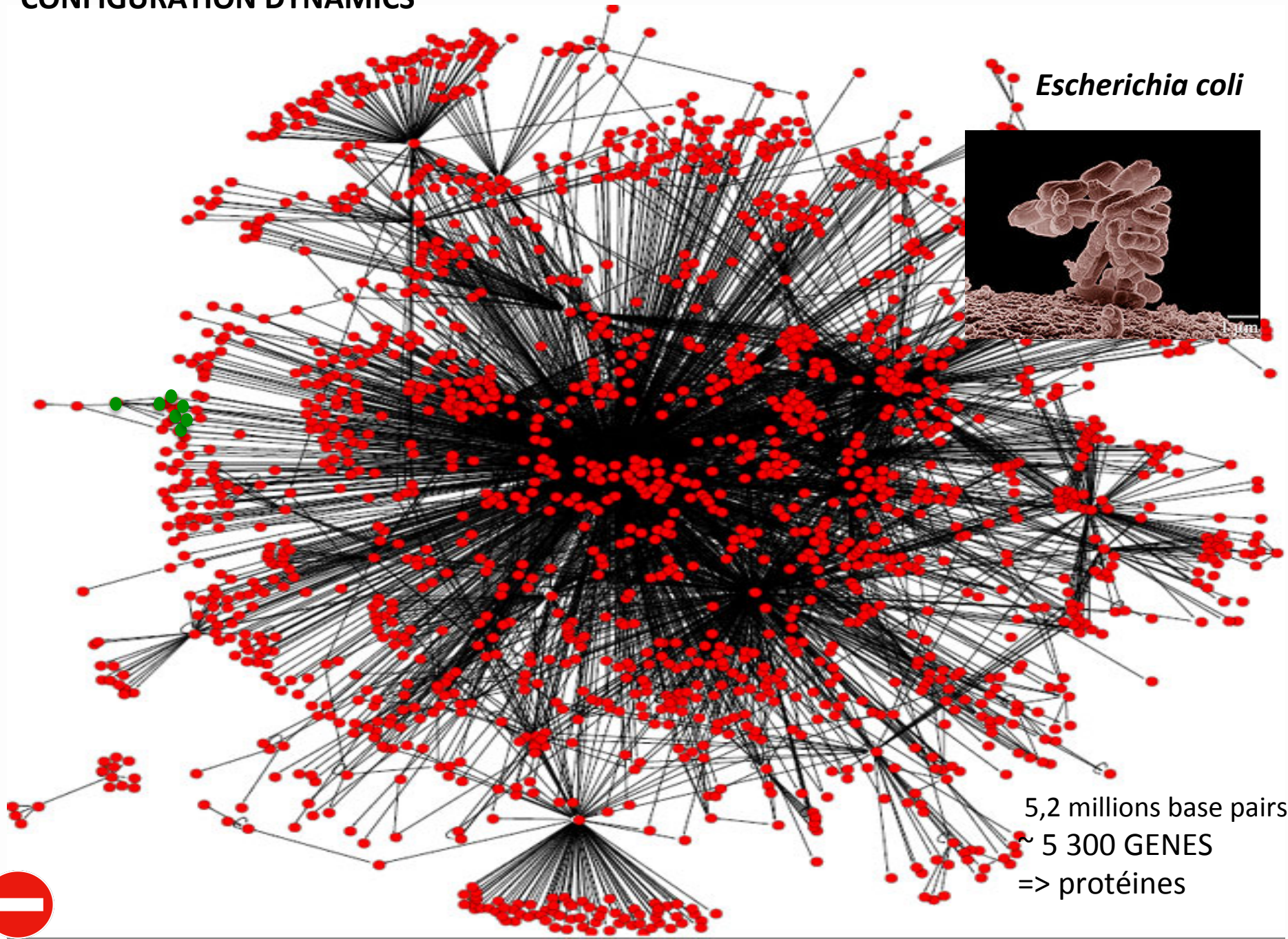
CONFIGURATION DYNAMICS



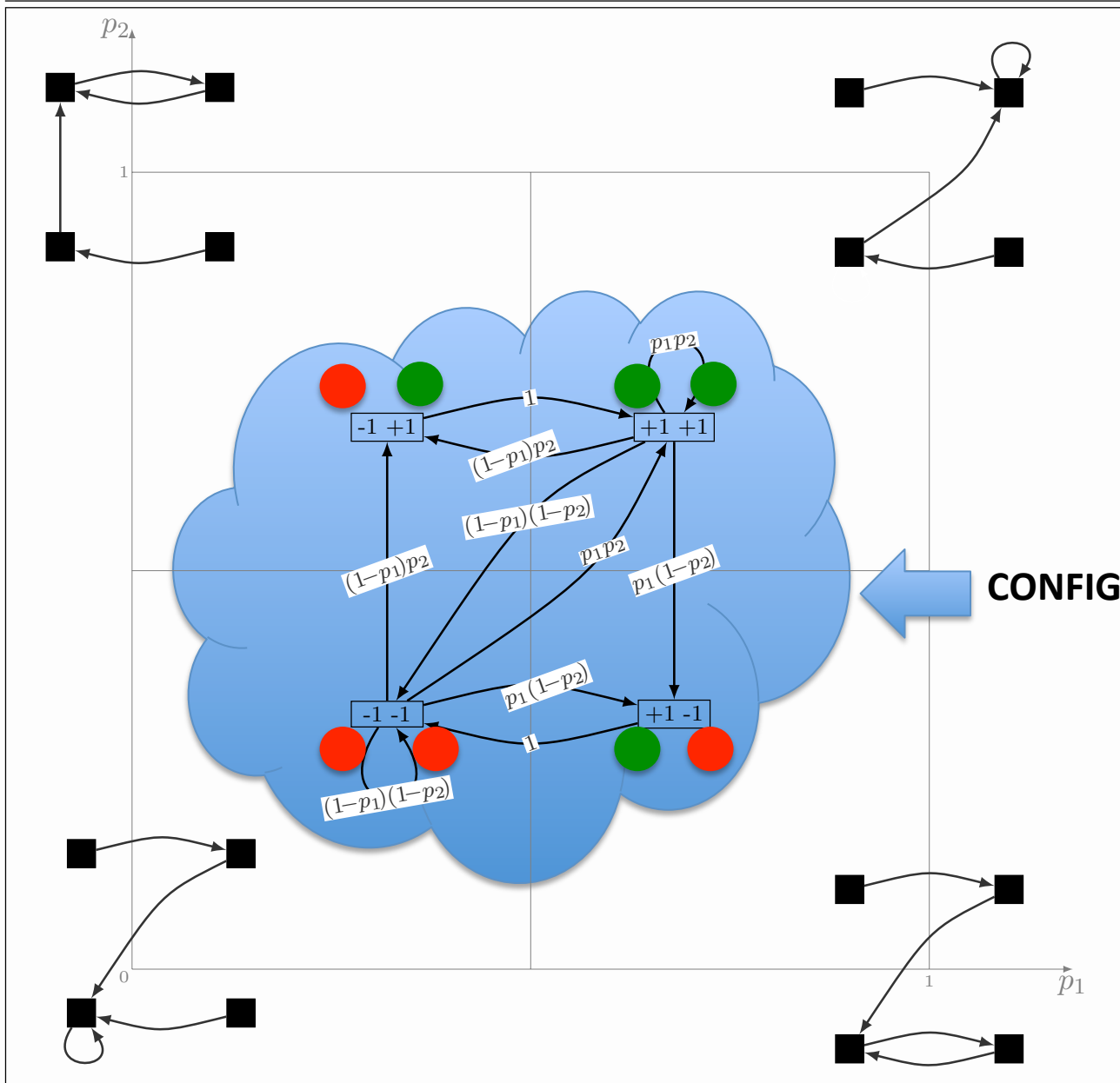
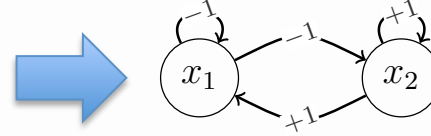
Escherichia coli

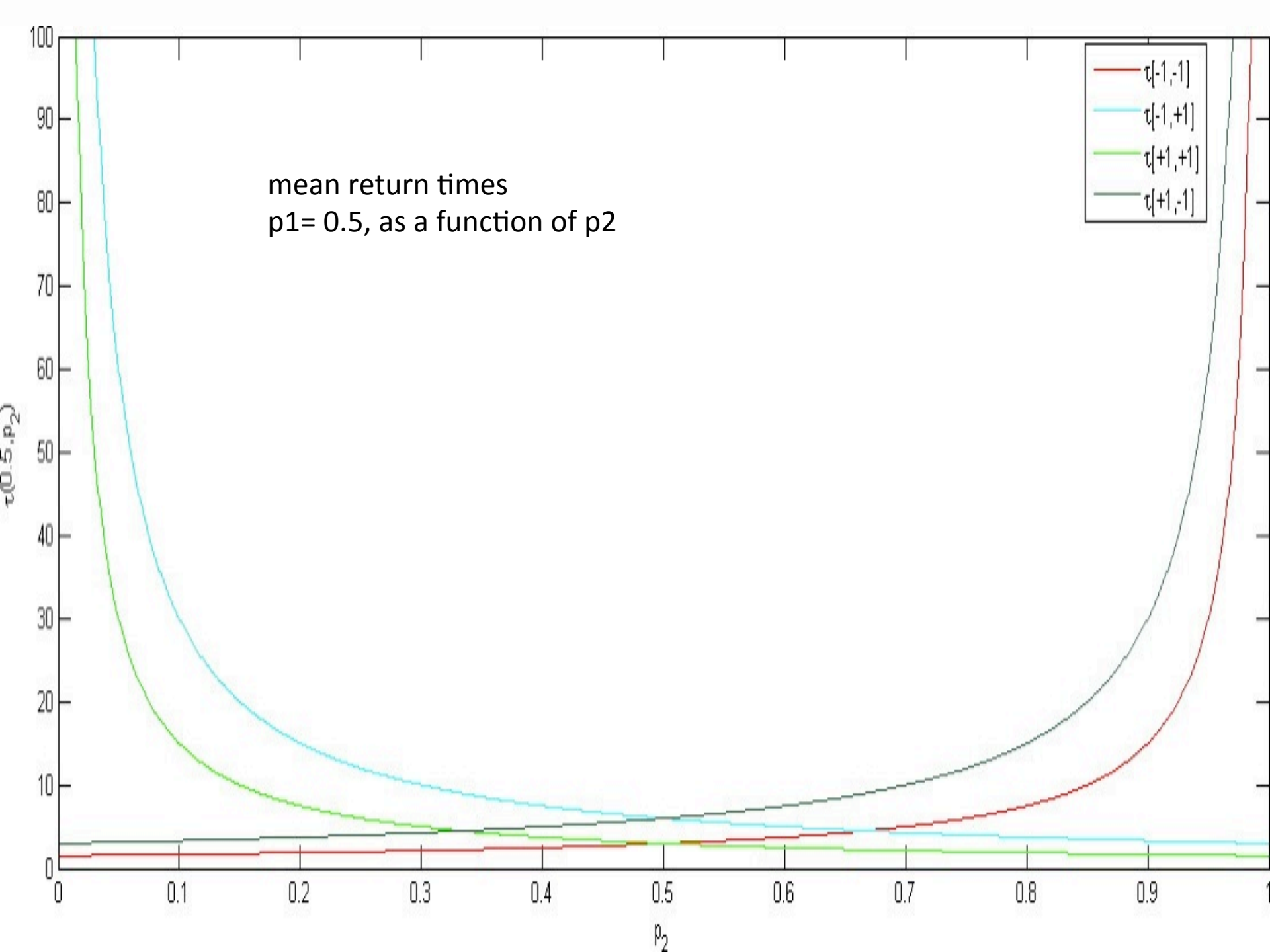
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CONFIGURATION DYNAMICS

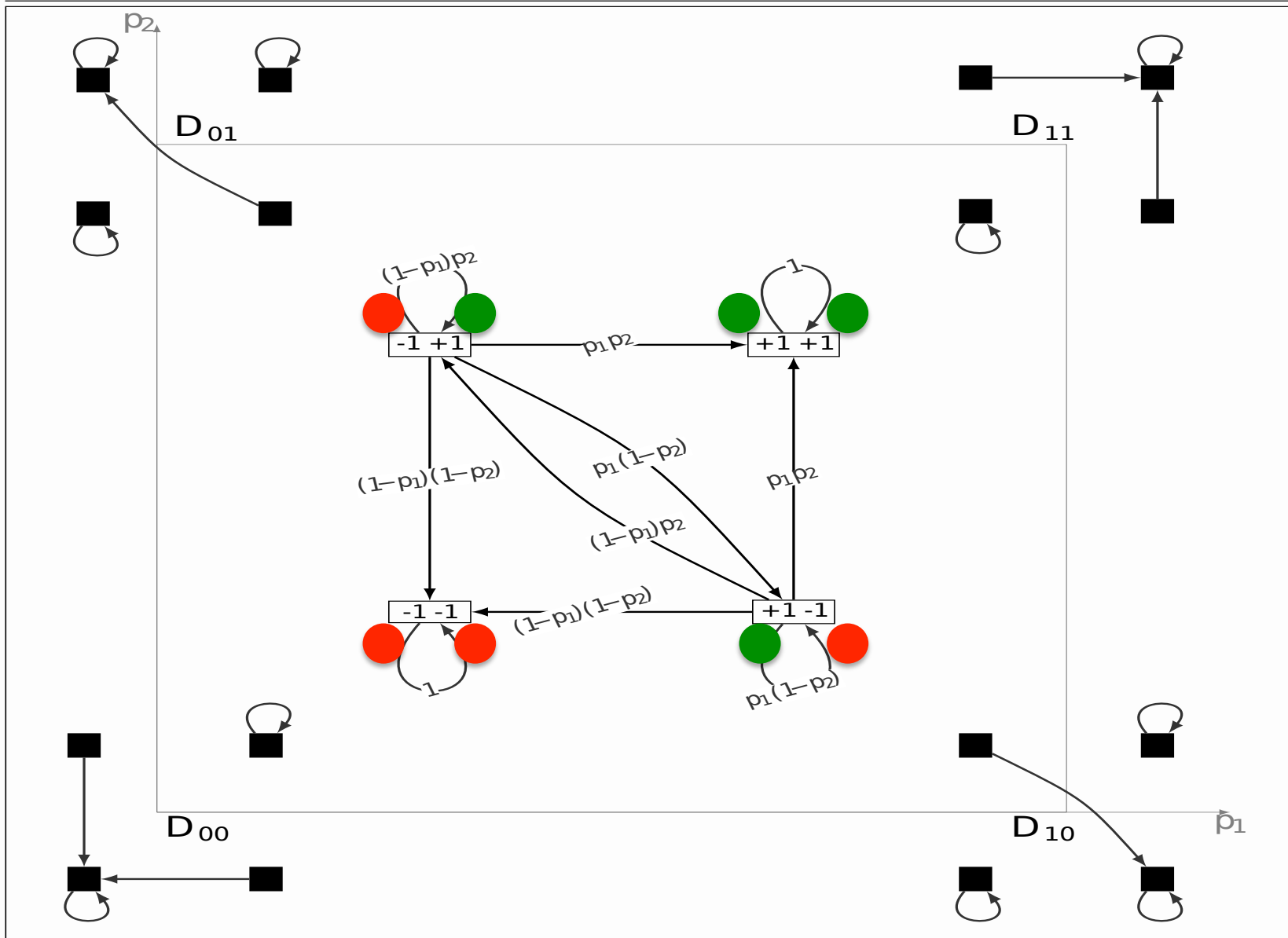
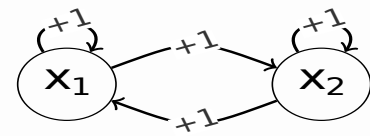


$[-1,+1] \oplus [+1,-1]$ (IP3)
2 GENE INTERACTION NETWORK

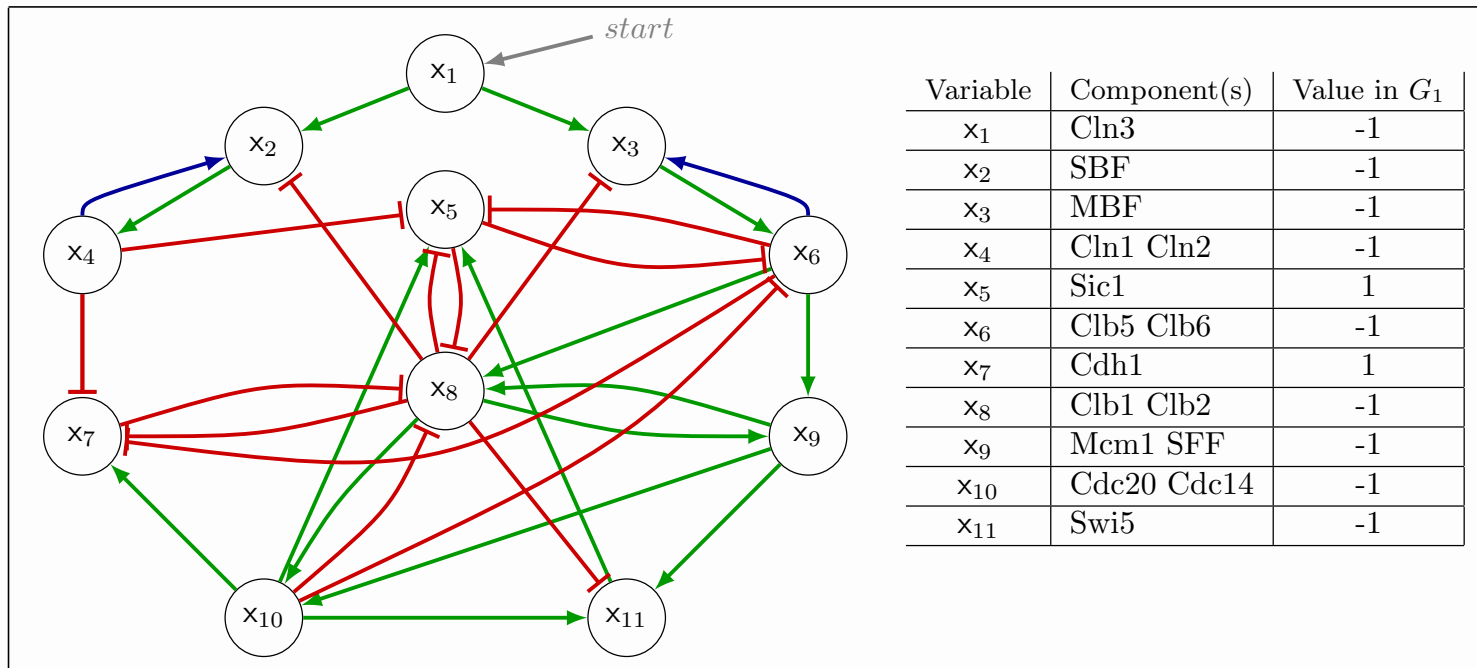




$[+1,+1]^\oplus$ $[+1,+1]$ (IP1)



EUKARYOTIC CELL CYCLE GENE NETWORK



The dynamics has a unique absorbing steady configuration: G_1
(adding the 2 blue interactions)

SOME IMPORTANT OPEN QUESTIONS

1) FROM INTERACTION GRAPH TO DYNAMICAL GRAPH:

Is it possible to directly study dynamical properties by inspecting the interaction network?

2) SEARCH OF ALL ATTRACTORS (ABSORBING SETS)

3) THE DYNAMICS OF A PROJECTION

4) SYSTEMATIC STUDY OF THE DYNAMICAL PROPERTIES OF MUTANTS

5) VARIABILITY FROM CELL TO CELL, INSIDE A POPULATION, etc.

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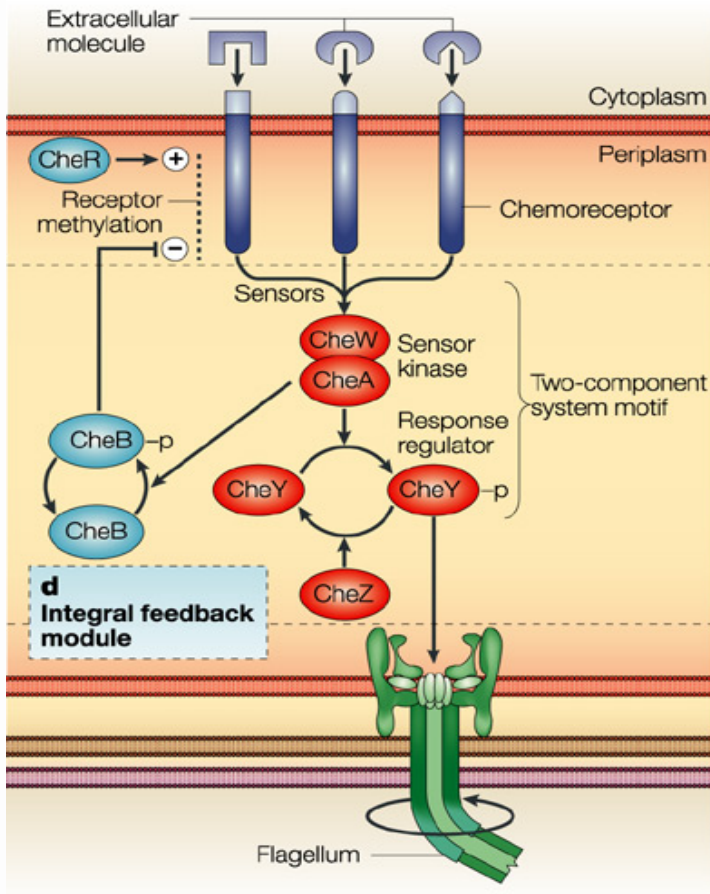
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A SMALL GENE NETWORK AT WORK: BACTERIA CHEMOTAXIS (*E. coli*)



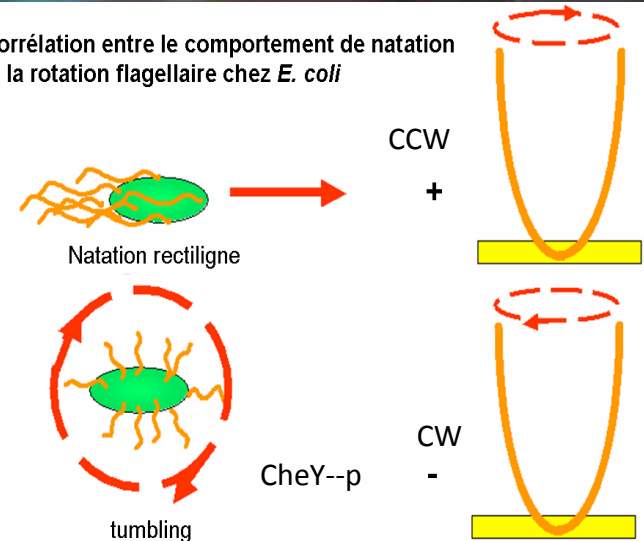
a Transmembrane chemosensor proteins form a **sensor module**

b Networked reactions between chemotaxis proteins create a **transduction module**

c Flagellum providing motive power for chemotaxis is the **actuator module**



Corrélation entre le comportement de natation et la rotation flagellaire chez *E. coli*



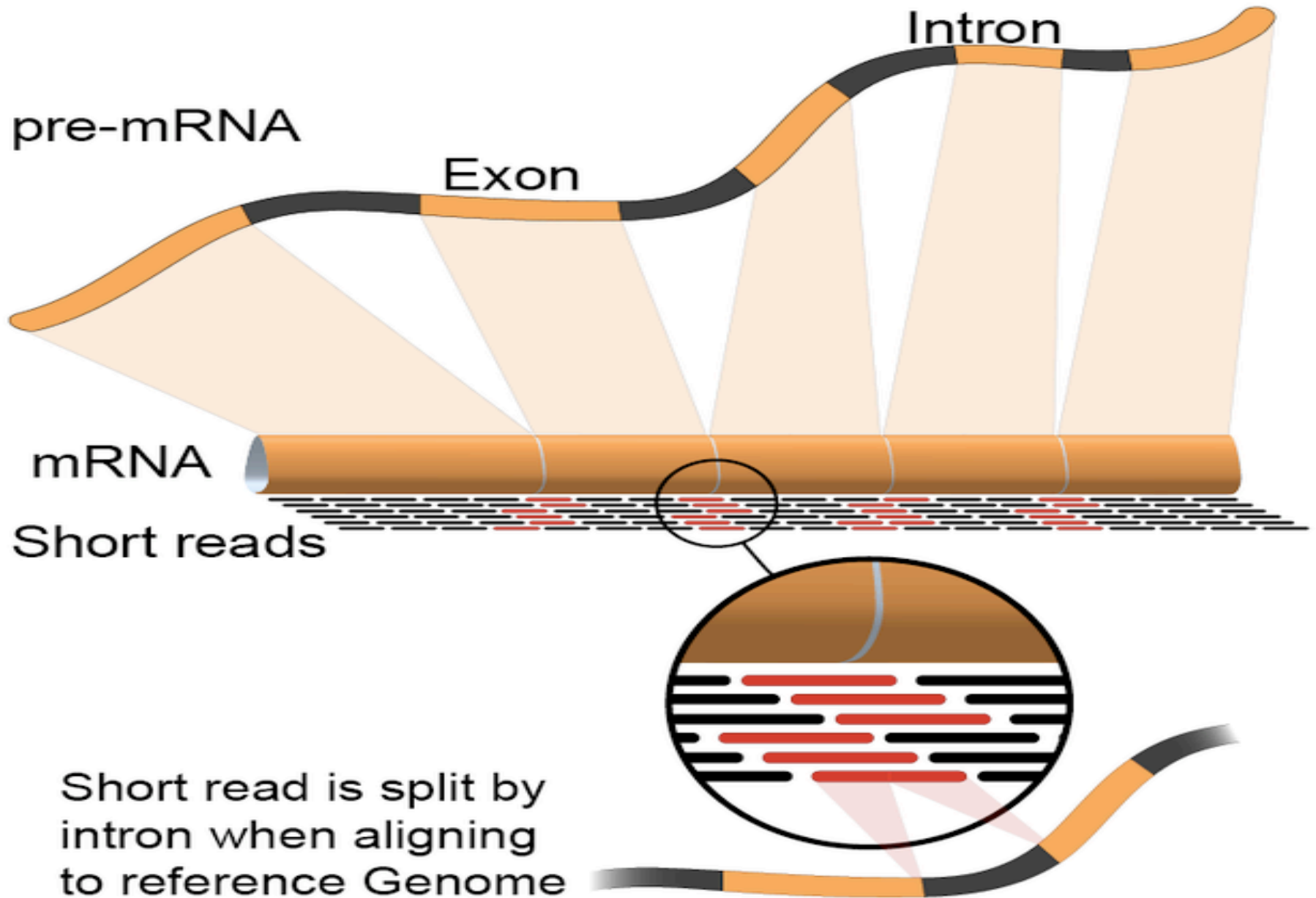
Nature Reviews | Genetics

*: sens inverse des aiguilles d'un montre, -: sens des aiguilles d'une montre

Attractants
Repellents
Flights (CCW) and Tumbles (CW)

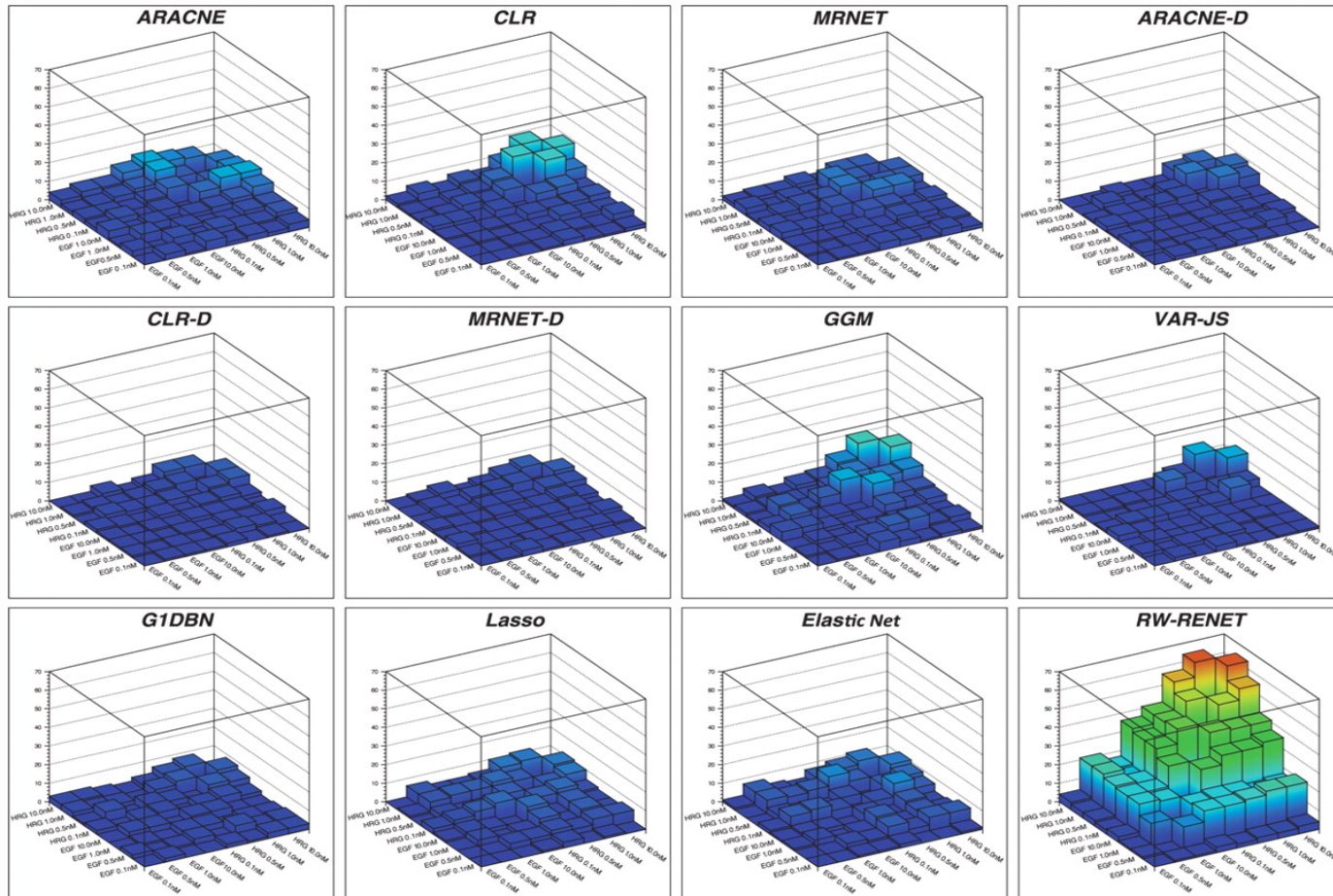
H.C. Berg, D.A. Brown, 1972, Nature 239: 500-504.

FROM READING SEQUENCES TO NETWORK INFERENCE



ON THE INCERTITUDE OF ALGORITHM INFERENCE

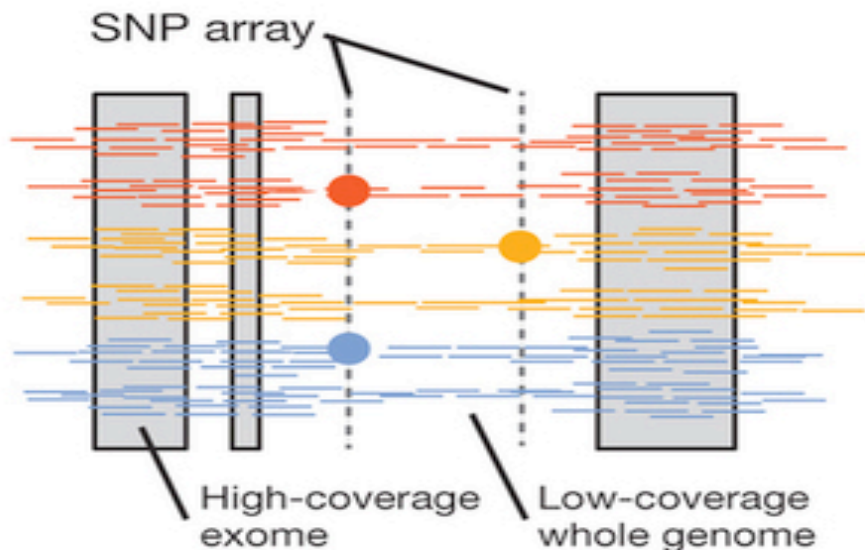
The 2D histograms showing how many edges were conserved between the inferred networks for each of the 12 inference algorithms (ARACNE, CLR, MRNET, ARACNE-D, CLR-D, MRNET-D, GGM, VAR with the James-Stein Shrinkage (VAR-JS), G1DBN, lasso, elastic net and RW-RENET).



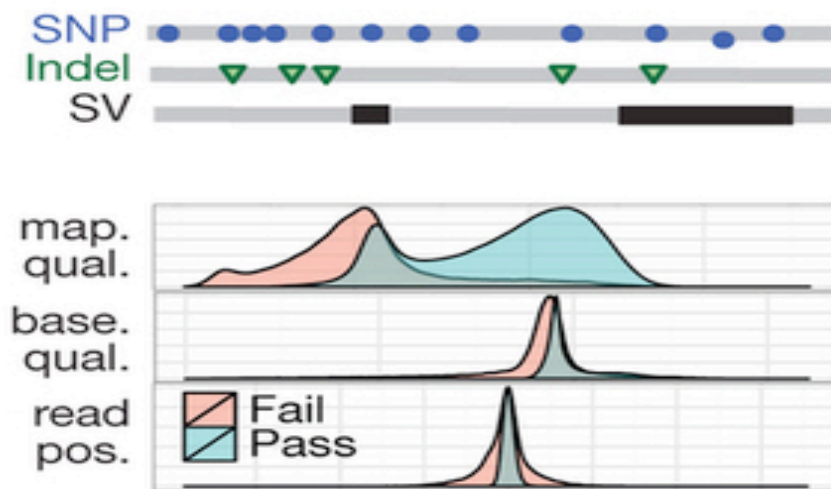
Shimamura T et al. *Bioinformatics* 2010;26:1064-1072

VARIABILITY

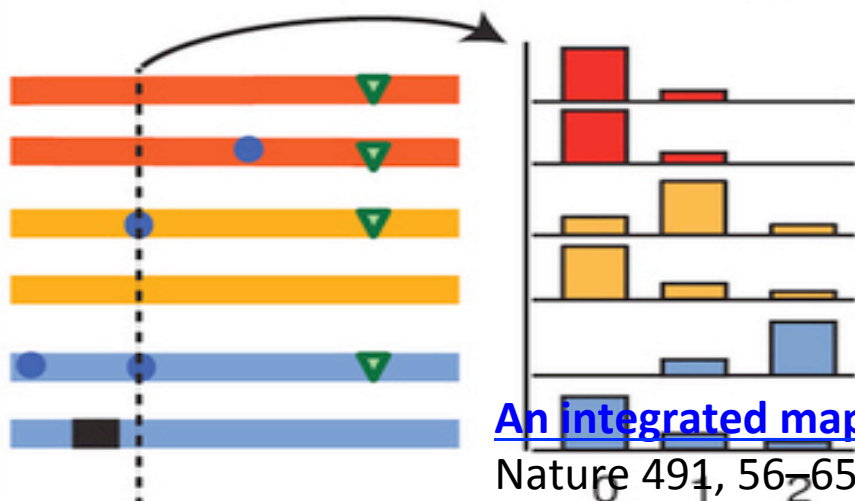
a Primary data
Sequencing, array genotyping



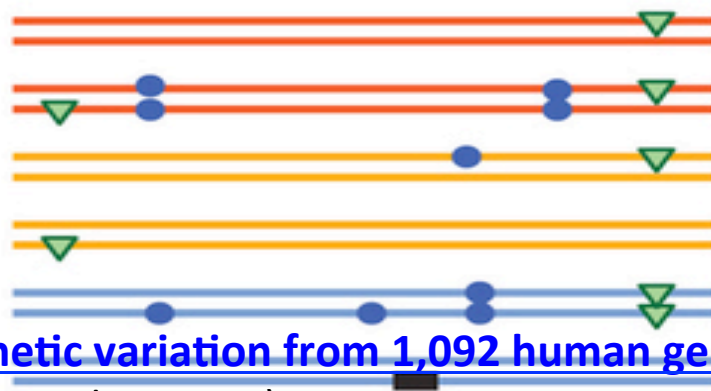
b Candidate variants and quality metrics
Read mapping, quality score recalibration



c Variant calls and genotype likelihoods
Variant calling, statistical filtering



d Integrated haplotypes
Probabilistic haplotype estimation

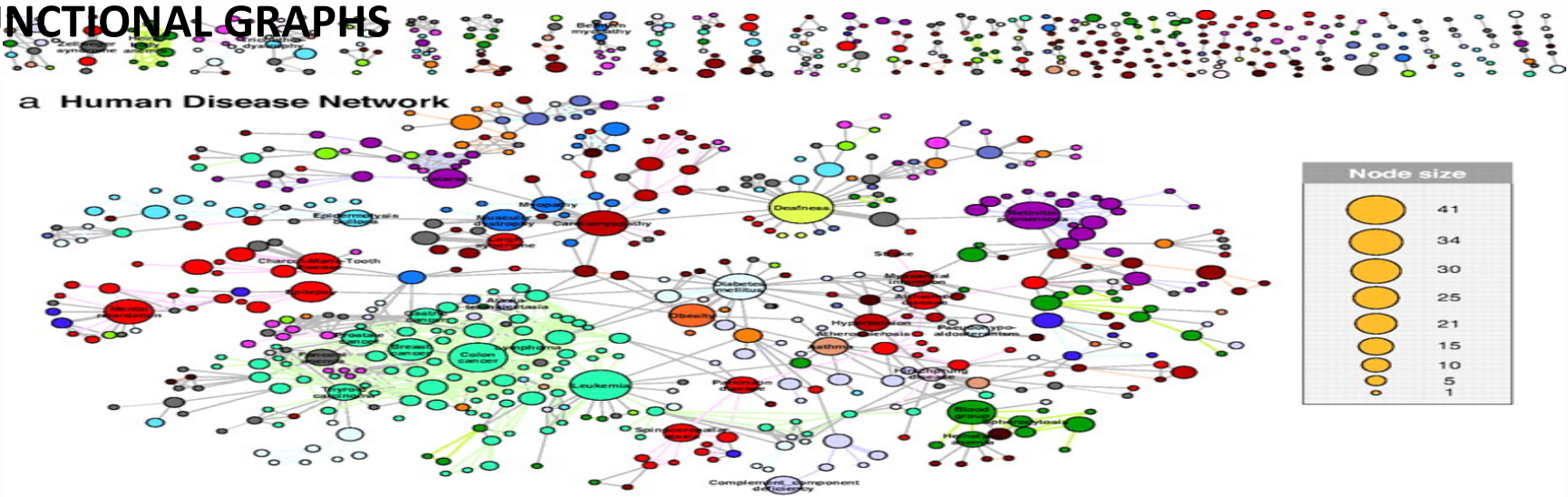


[An integrated map of genetic variation from 1,092 human genomes](#)

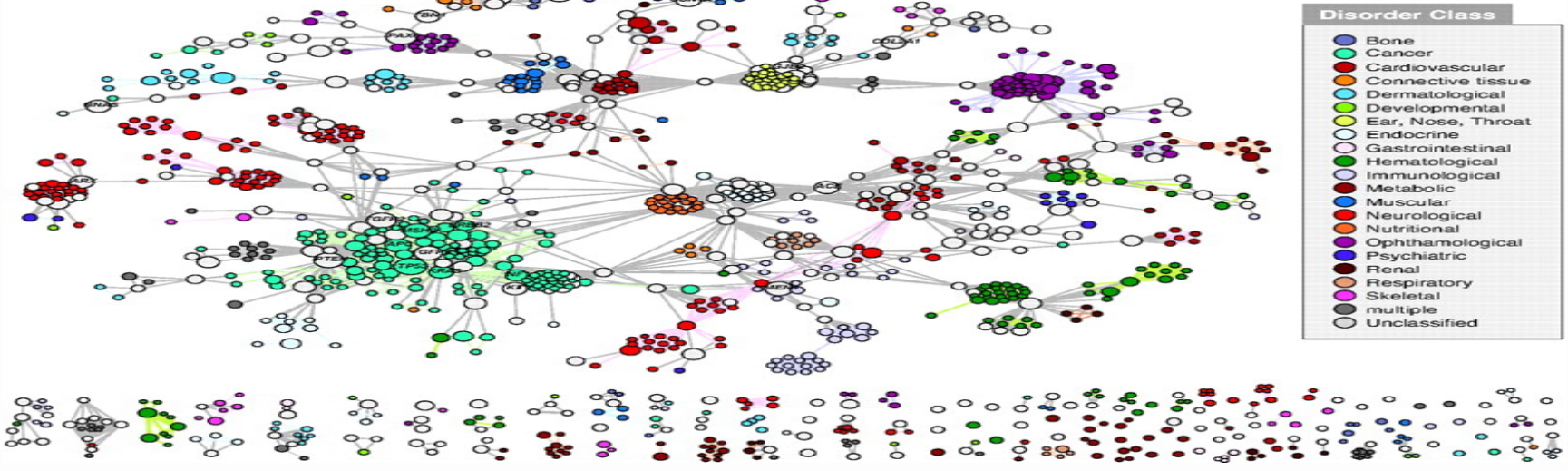
Nature 491, 56–65 (01 November 2012)

FUNCTIONAL GRAPHS

a Human Disease Network

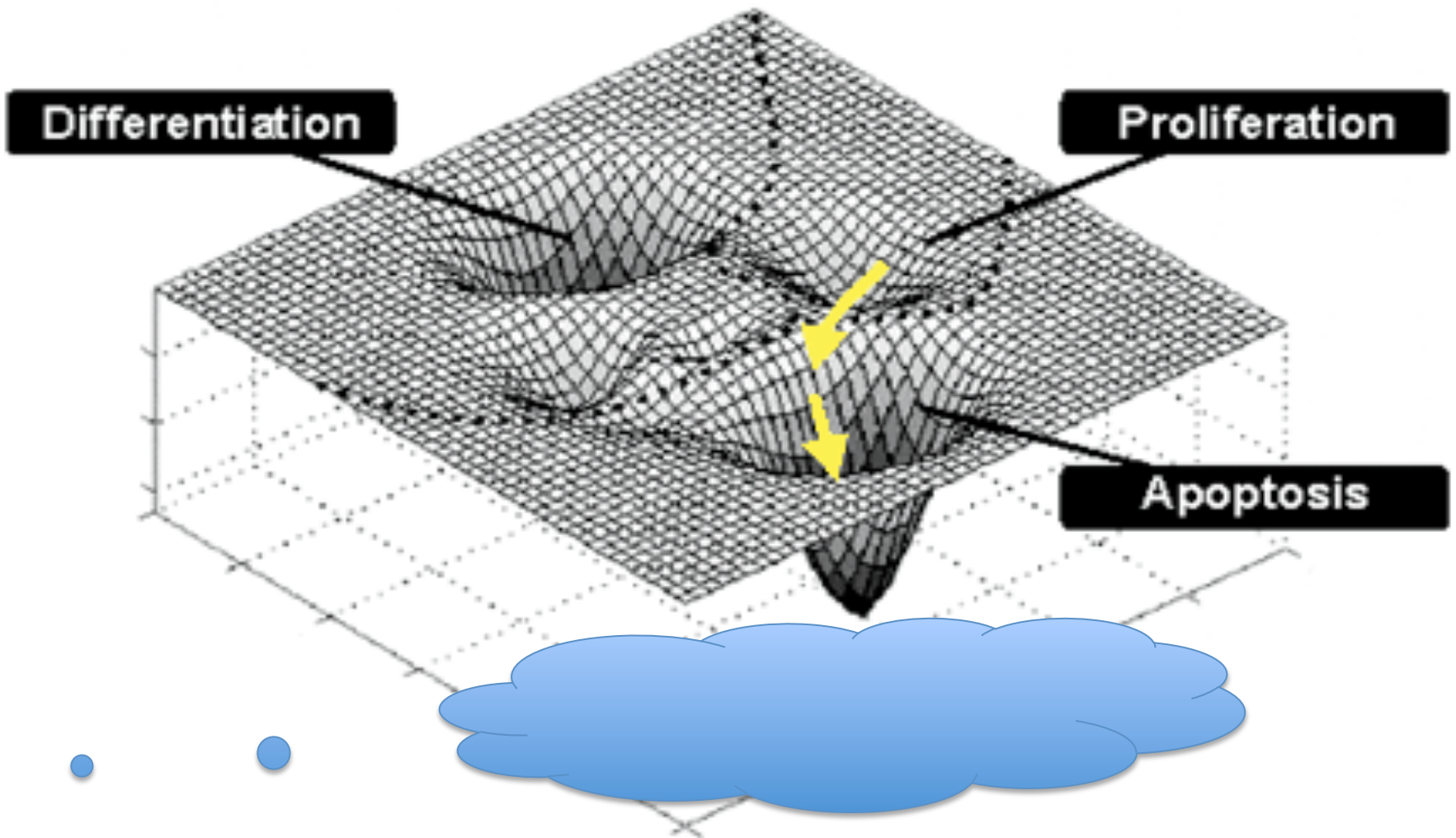


b Disease Gene Network



Goh K et al. PNAS 2007;104:8685-8690

PNAS



Attractor landscape representation of cell fate determination. A hypothetical 'potential landscape' that represents the n-dimensional state space compressed into two dimensions (XY) for visualization purposes.

FINAL QUESTIONS:

1) When is gene regulation needed or benefit for a cell (or an organism)?

Under rapid changes of the environment. [E. Hartl and all, 84]. Other situations ?

2) Adaptability versus optimality [G. F. Gause , 34 ...] : why modular organization of the network won [L. L. Hartwell and all , 99 ...] ?

-multi needs/task and time change in the environment + slow structural change capabilities call for modularity instead of optimality. Fitness function not always exists or a good definition is not known.

-.

**3) What are the known mechanisms of robustness in transcription regulation?
(against intrinsic stochasticity and extrinsic noise)**

(a) Integral feedback (negative feedback proportional to time integral of the difference of the actual level to the goal. [N. Barkai, S. Leibler, 97]

(b) Kinetic proofreading (time-delay of signal, small error rate of recognition) [J. Hopfield, 74]

(c) Self-enhanced degradation (non-linear diffusion) minimize errors in pattern location [A. Eldar and all, 02]

(d) When stochasticity is a most [ex. T-cell allelic exclusion]

4) How the description of a specific level can be integrated in the next level ?

Module to network.

Single cell to cell population.